

Campylobacter data from a Turkish University hospital laboratory

Yesim Gurol¹, Zehra Kipritçi¹, Suat Biçer², Ibrahim Çağatay Acuner¹, Ayça Vitrinel², Gülden Çelik¹

(1) Medical Microbiology Laboratory, (2) Department of Child Health and Diseases, Yeditepe University Hospital, Istanbul, Turkey.

To the Editor,

Campylobacter spp. are primarily zoonotic, with a variety of animals implicated as reservoirs for human infection. Campylobacteriosis is common in both developed and nondeveloped countries. Definitive diagnosis requires isolation of bacterium in culture (i.e. “gold standard”). Due to underdiagnosis and underreporting, actual incidence of infection in any country is substantially greater than reported incidence (1). Here, we report laboratory-based surveillance data collected between 2009 and 2012 in Yeditepe University Hospital (JCI Accredited, 2010-2013, Istanbul, Turkey) Medical Laboratories (ISO-EN 15189 Accredited by DACH, 2009-2014), Microbiology Section, Bacteriology Sub-disciplinary Laboratory. Routine laboratory workflow for faecal specimen cultures included algorithms for isolation and identification of major bacterial pathogens for enteric infections. National Laboratory Surveillance Network for Enteric Pathogens (Microbiology Reference Laboratories, Turkish Public Health Institute, Ankara) requires laboratory-based direct reporting of *Salmonella* spp., *Shigella* spp., Enterohemorrhagic *Escherichia coli* (EHEC) (i.e O157 and non-O157) and *Campylobacter* spp. isolates. Among 3510 faeces cultures, *Salmonella* spp. (n = 135), *Shigella* spp. (n = 2), *Campylobacter* spp. (n = 142) and *Aeromonas* spp. (n = 4) isolates were detected. Out of 142 *Campylobacter* spp. isolates, the distribution was as follows : *C. jejuni* (115), *C. upsaliensis* (13), *C. coli* (6) and other species (8). Faecal samples were cultured in *Campylobacter*-BAP medium (Salubris, Turkey) and incubated under microaerophilic conditions (CampyGen, Oxoid, UK) at 42°C for 48 hours. Suspected colonies were examined by Gram stain and evaluated for oxidase and catalase positivity. Subsequently, suspected isolates were also characterized with *Campylobacter* latex agglutination test (Dryspot, Oxoid, UK) and API CAMPY biochemical identification system (bioMérieux, France). Antimicrobial susceptibility of isolates against erythromycin was also tested with API CAMPY system (bioMérieux, France). The distribution of isolates, demographic information and seasonal epidemiology in patients with Campylobacteriosis are shown (Table 1, Fig. 1). In faecal cultures, *Campylobacter* spp. were the most frequently isolated bacterial enteric pathogen, and relative frequency of Campylobacteriosis was 50%. Most studies report that *Campylobacter* infections are typically seen in summer season whereas our data

show that infections occurred most frequently in March, May, July and August (2,3,4). Mostly Campylobacteriosis cases are managed by fluid therapy, however antibiotic therapy may be required in severe cases. Macrolide resistance rate is lower than 5% in most countries while increasing in some areas of the world due to ribosomal mutations. Fluoroquinolone resistance rates are between 10% and 20% in most Europe countries, however higher rates up to 50% may be observed in many developing countries (5). In antimicrobial susceptibility testing of *Campylobacter* spp., disk diffusion, broth microdilution, agar dilution or Etest methods may be used. CLSI suggests disk diffusion and broth microdilution methods as standardized antimicrobial susceptibility tests in *Campylobacter* spp (6). In our routine laboratory workflow, erythromycin susceptibility is tested with API CAMPY System, bioMérieux, France. In this study period, erythromycin resistance rate was 7%. Reported fluoroquinolone and macrolide resistance rates in *Campylobacter* spp. may considerably vary between countries. Therefore, laboratory-based surveillance is an important tool for early detection of changes in local epidemiology of antimicrobial resistance in *Campylobacter* spp.

Table 1. — Distribution of isolates among age

	0-5 age (n = 52)	6-14 age (n = 42)	15-34 age (n = 27)	> 35 age (n = 17)
<i>Campylobacter jejuni</i>	82.7%	83.3%	77.8%	70.6%
<i>Campylobacter</i> spp.	5.8%	7.2%	3.7%	5.9%
<i>Campylobacter coli</i>	1.9%	2.4%	7.4%	11.7%
<i>Campylobacter upsaliensis</i>	9.6%	7.1%	11.1%	11.8%

Correspondence to: Yesim Gurol, M.D., Assoc. Prof., Medical Microbiology Laboratory, Yeditepe University Hospital, Devlet Yolu Ankara, cad.No : 102-104 Kozyatagi, Istanbul, Turkey. E-mail : yesimg@yeditepe.edu.tr

Submission date : 23/08/2012

Acceptance date : 12/10/2012

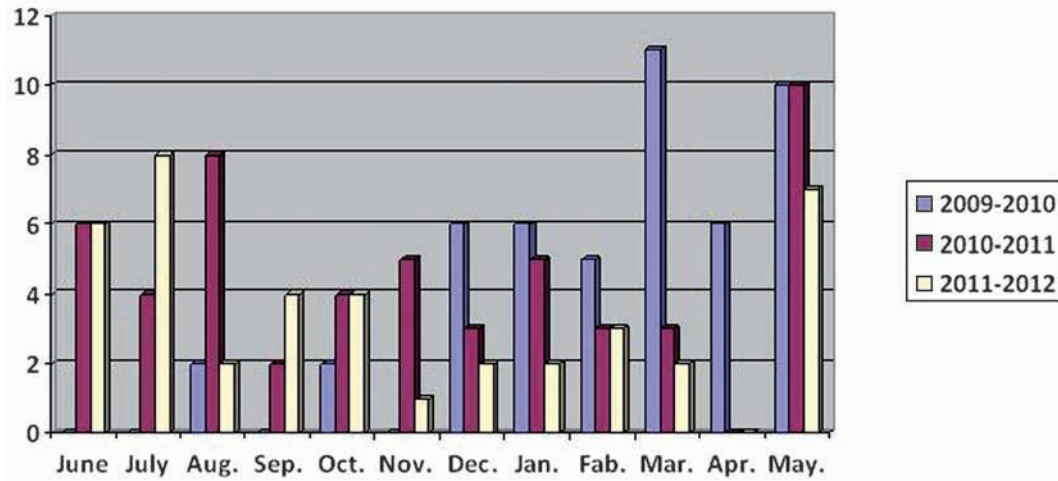


Fig. 1. — Figure 1 shows the seasonal epidemiology in our patients with Campylobacteriosis occurring most frequently in March, May, July and August.

References

1. FITZGERALD C., NACHAMKIN I. Campylobacter and Arcobacter. In: Manual of Clinical Microbiology. Volume 1. 9th ed. Washington, ASM Press, 2007 : 933.
2. ÖZTÜRK R., MIDILLI K., OKTAY K. Çocuk ve erişkin yaş grubu sürgün olgularında C. jejuni ve C. coli sıklığının araştırılması. *Türk. Mikrobiyoloji Cem. Der.*, 1994, **24** : 42-45.
3. YILDIRIM M.S., FAZLI Ş.A. Kayseri ve yöresinde bakteriyolojik kültür için gönderilen dışkı örneklerinde Campylobacter'lerin izolasyonu ve identifikasyonu. *İnfeksiyon Derg.*, 1998, **12** (3) : 317-322.
4. ÖNGEN B., NAZIK H., KAYA I. Rutin dışkı kültürlerinde üretilen Campylobacter türleri ve antibiyotik duyarlılıkları : 5 yıllık sonuçların değerlendirilmesi. *Ankem. Derg.*, 2007, (1) : 37-41.
5. SAENZ Y., ZARAZAGA M., LANTERO M., GASTANARES M.J., BAQUERO F., TORRES C. Antibiotic resistance in Campylobacter strains isolated from animals, foods, and humans in Spain in 1997-1998. *Antimicrob. Agents Chemother.*, 2000, **44** : 267-271.
6. YILMAZ AKYÖN Y. Campylobacter türleri ve Helicobacter pylori : Antimikrobiyal Duyarlılık Testleri. *Gülhane Mikrobiyoloji Günleri Özet Kitabı*, 2010 : 122-123.